

Title: HYDROGEN STORAGE MATERIAL
BASED ON A MULTILAYERED CORE/SHELL
STRUCTURE

Inventor's Name: Peter C. EKLUND, et al.

Application No.: New Application

Docket No.: 025756-00003

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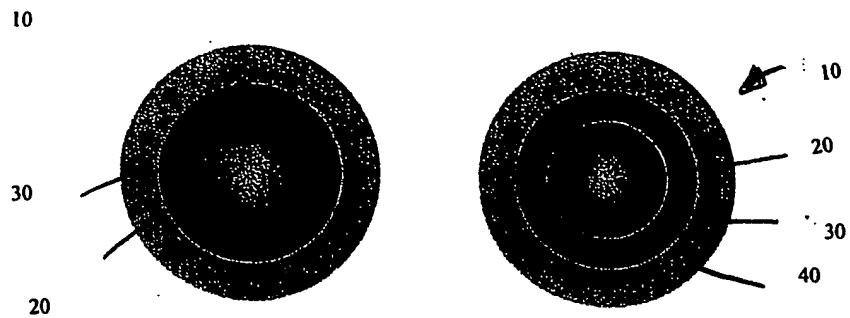


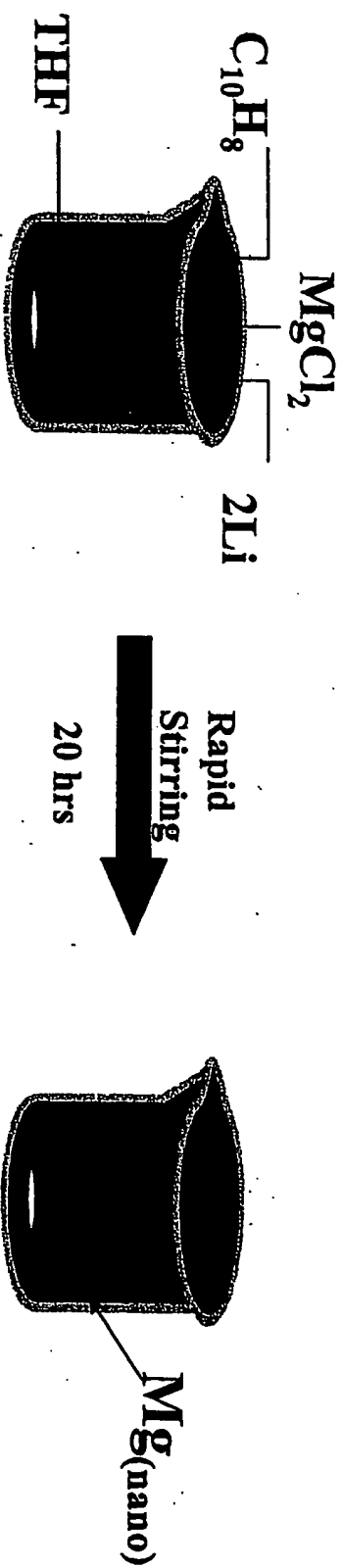
FIG. 1A

FIG. 1B

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Mg Synthesis

1. Synthesis (performed in an Ar Glove Box)



2. Remove THF with dissolved by-products

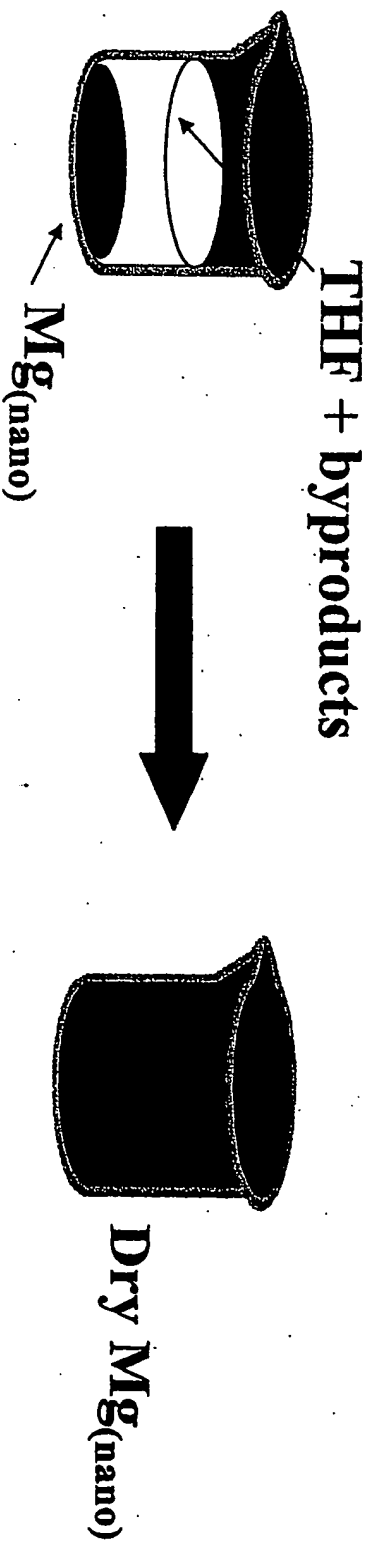


FIG. 2

Synthesized Mg

FIG. 3

Sample	Amount Made	XRD crystallite size (Scherrer equation)*
Mg 1-2	0.198g	23nm
Mg 1-10	0.4g	Used in Pd capping
Mg 1-13	0.4g	37nm
Mg 1-18	0.2g	34nm
Mg 1-41	0.2g	Used for Co capping
Mg 1-51	0.4g	33nm
Mg 1-57a	0.5g	25nm
Mg 1-57b	0.5g	Still in dry box
Mg 1-57c	0.5g	Almost amorphous

$$*t = [((0.9)(0.154))/((\beta)(\cos \theta))]$$

t = crystallite size in nm

β = full width half max

θ = Bragg angle

Klug and Alexander, 1950

As Synthesized Mg XRD

XRD of Mg 1-13 sample matches that of an indexed Mg pattern
Material is coated with amorphous sp² carbon allowing it to be stable in air

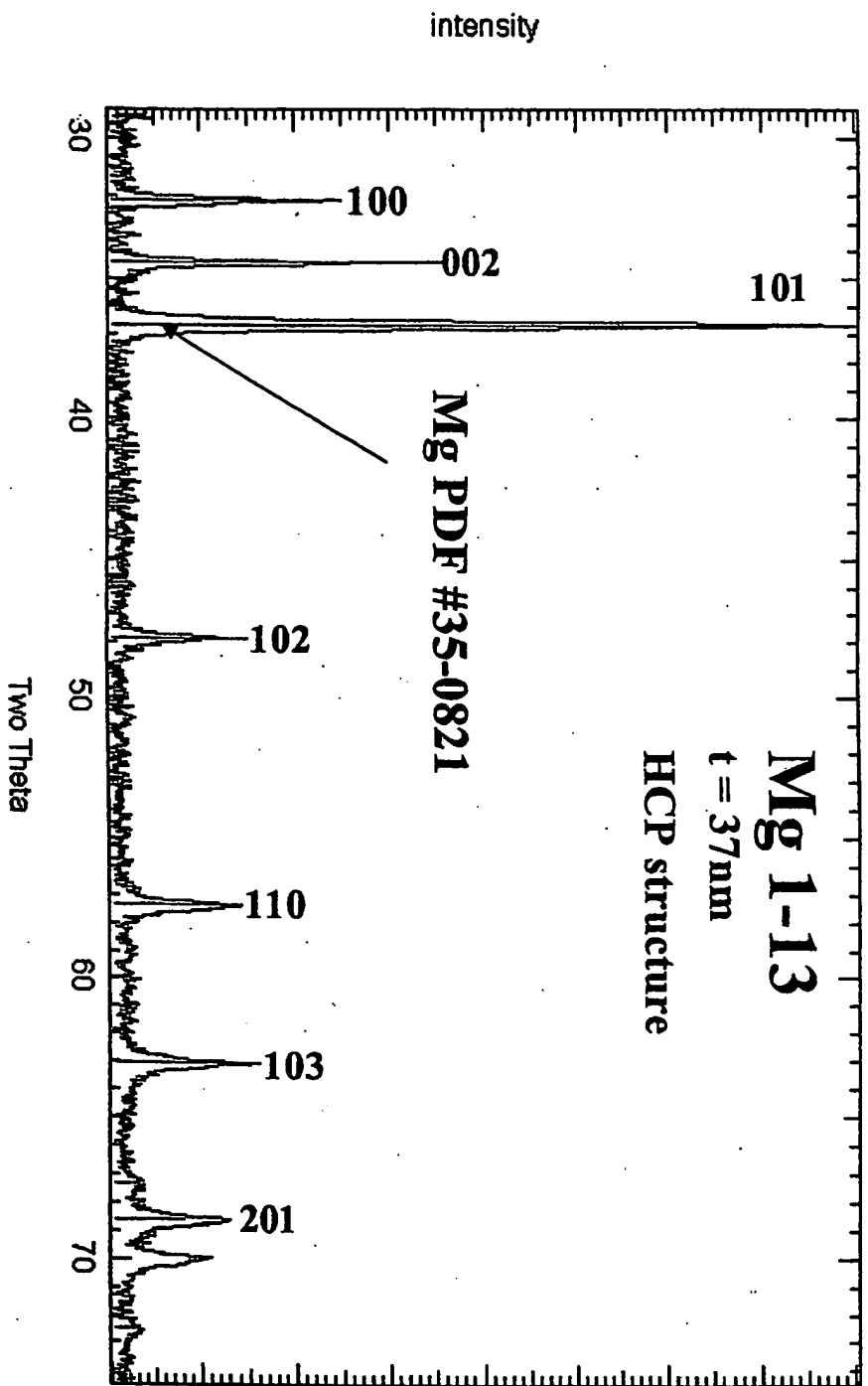


FIG. 4

Still Mg Metal!

Mg 1-13 sample was stable in atmospheric conditions for 3 months

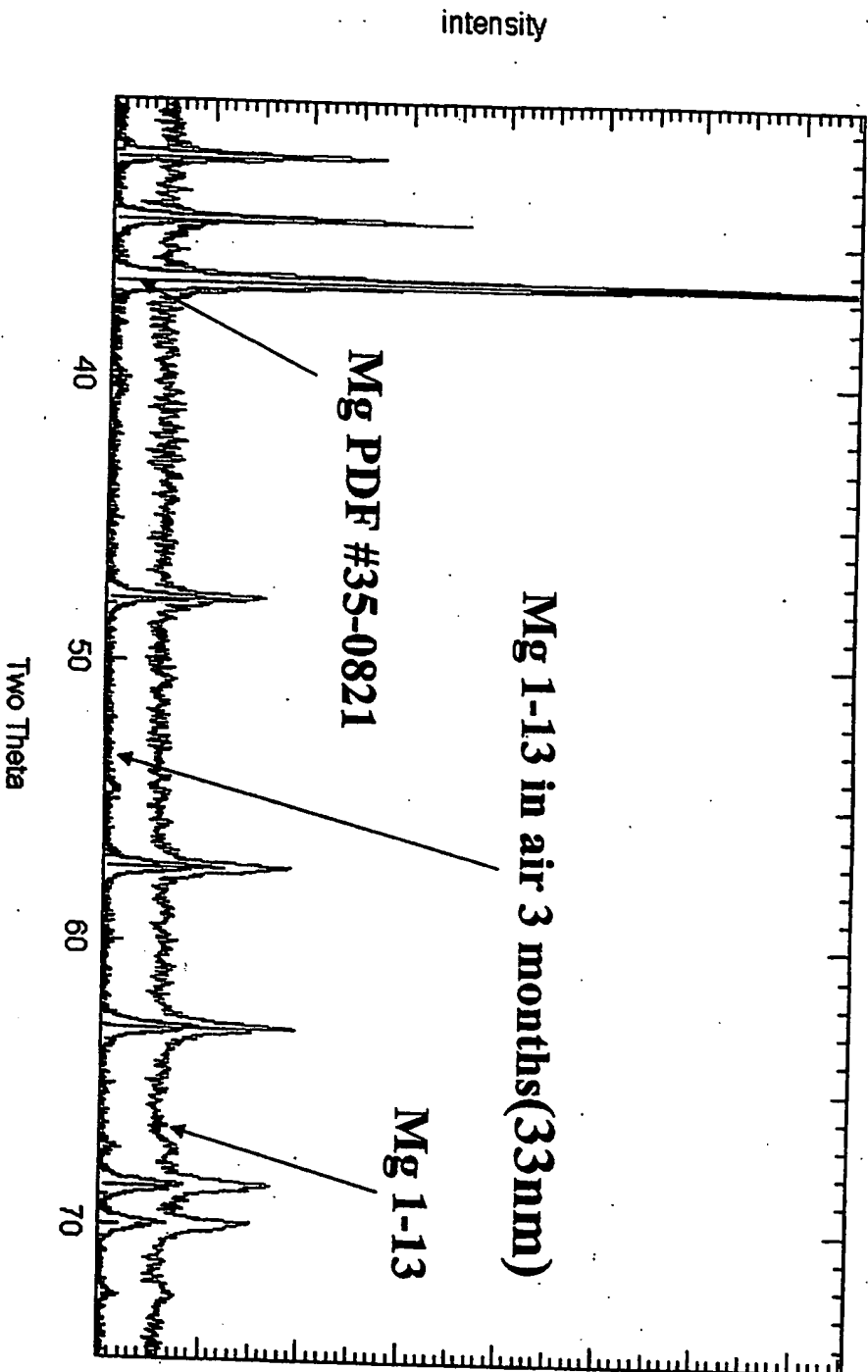


FIG. 5

Phase Stability in Water

After exposure to water for ~48 hours \Rightarrow

Mg 1-13 turned white with XRD giving $\text{Mg}(\text{OH})_2$

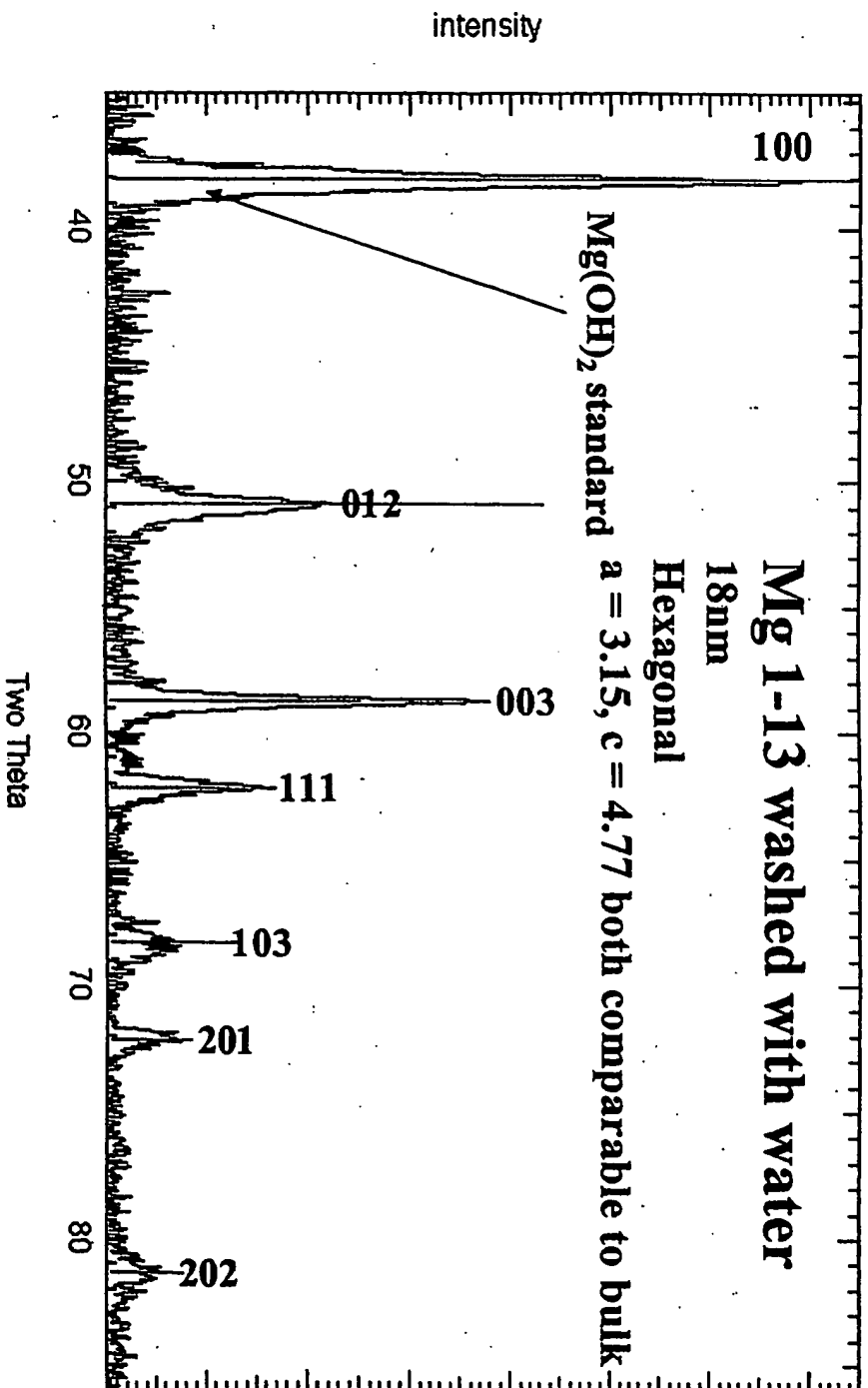


FIG. 6

Pd coated Mg Synthesis

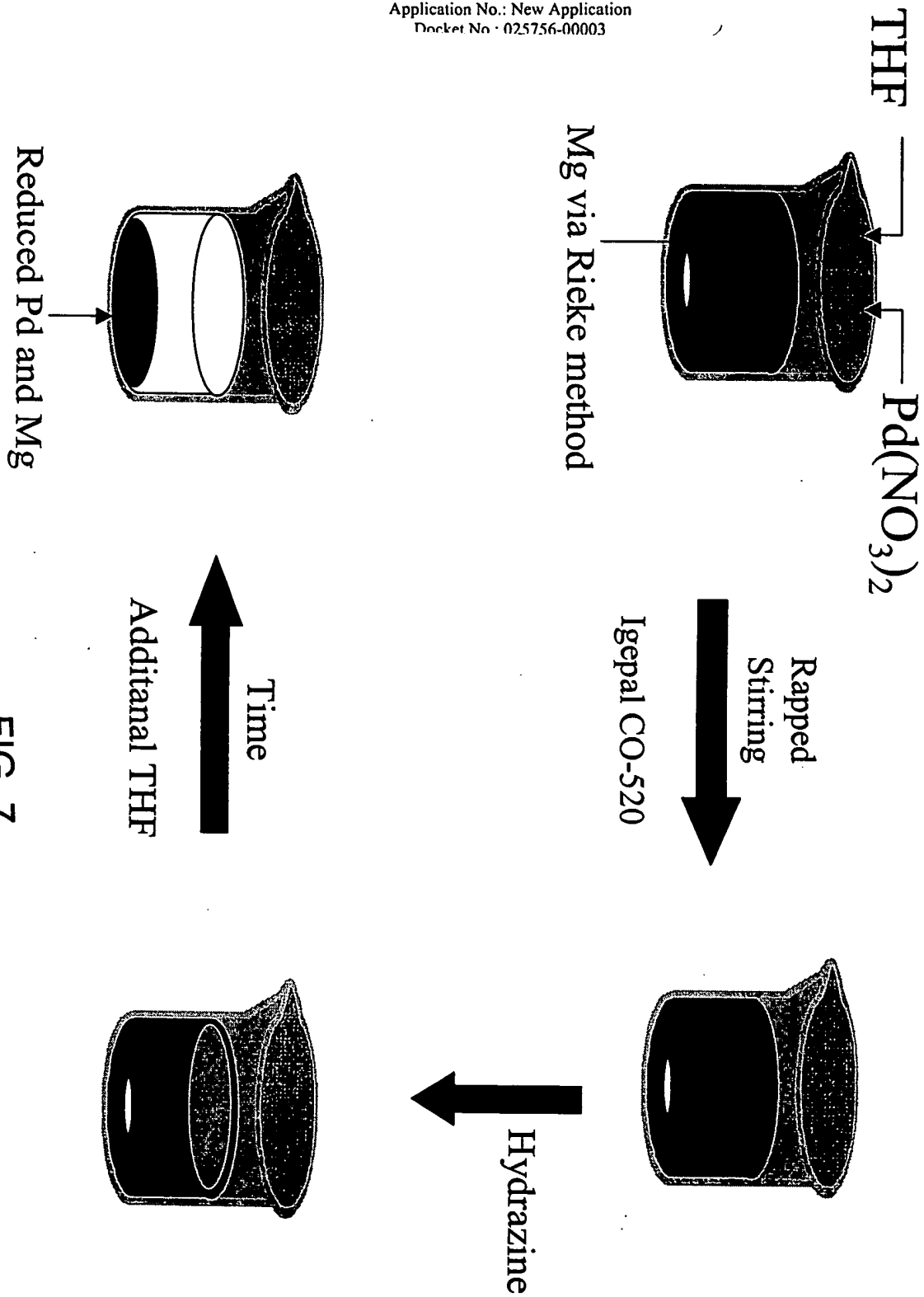


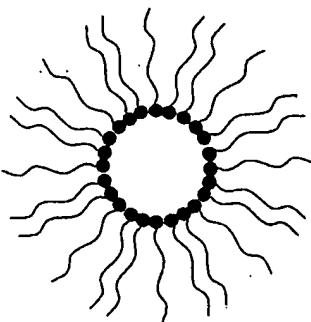
FIG. 7

Spherical nPd

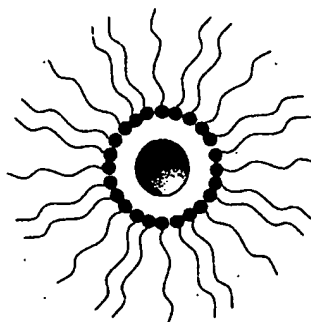
1) Micelle Formation

Igepal™

cyclohexane



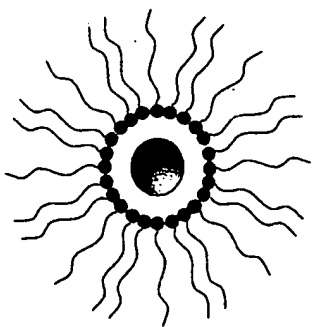
0.04 M Pd in H₂O



2) Reduction of Pd²⁺ using Hydrazine Hydrate (H₂NNH₂•xH₂O)



3) Break the Micelle and collect the Pd with ethanol



Ethanol



Pd
nano

FIG. 8

FIG. 9 Controlling The Size

The size of the Pd spherical particle is dependent on the mole% ratio of surfactant (Igapal) to water often referred to as the R value.

$$R = [\text{water}]/[\text{surfactant}]$$

The larger the R value the larger the radius of the particle

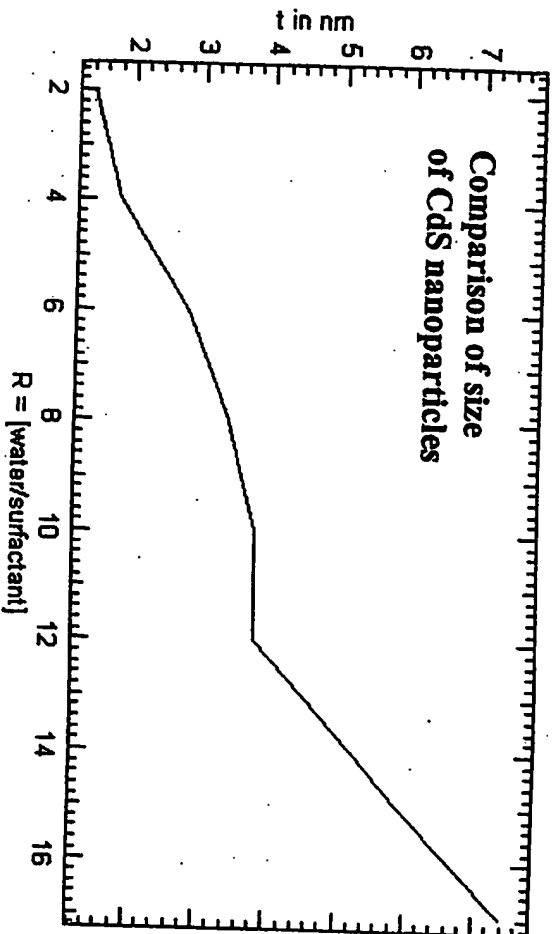


FIG. 10 Spherical Pd Synthesized to Date

Sample	R value	Amount made	Particle size*
Pd 1-62	8	1.3mg	
Pd 1-67	6	4mg	
Pd 1-68-1	1	2mg	
Pd 1-68-2	2	4mg	
Pd 1-69-1	1	.67mg	
Pd 1-68-2	2	1.35mg	
Pd 1-69-6	6	12.3mg	
Pd 1-69-8	8	16.32mg	
Pd 1-70-7	7	19mg	
Pd 1-70-9	9	30.5mg	~9nm
Pd 1-71	9	61mg	
Pd 1-73	5	63.6mg	
Pd 1-74-3	3	61mg	
Pd 1-74-7	7	71mg	
Pd 1-91-6	6	81.4mg	5nm
Pd 1-91-8	8	108.5mg	4nm
Pd 1-96	7	399mg	
Pd 1-100	25	84.8mg	6nm

*By Scherrer equation

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DF TEM Spherical Pd

Pd 1-91 R=8; t=4nm

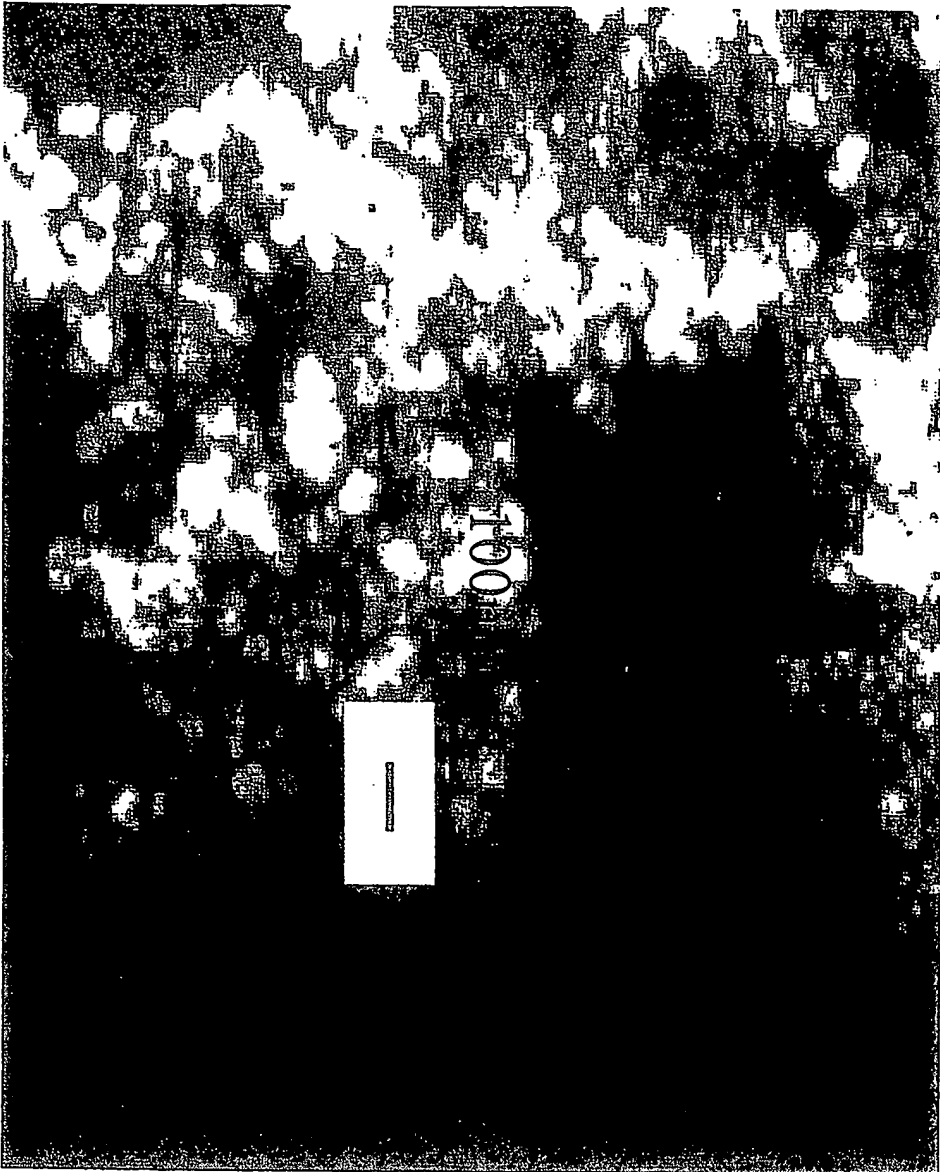


FIG. 11

XRD Of Spherical Pd

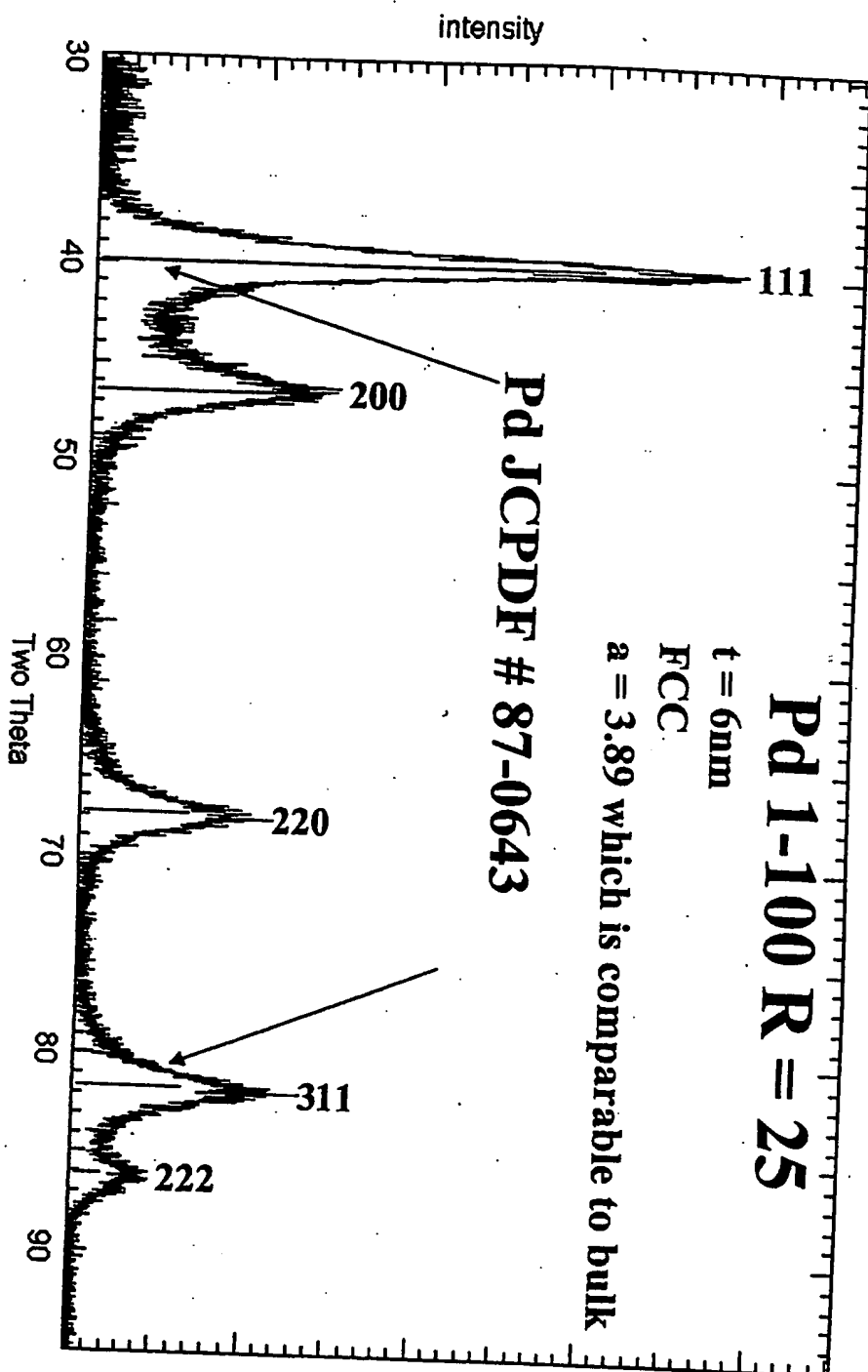
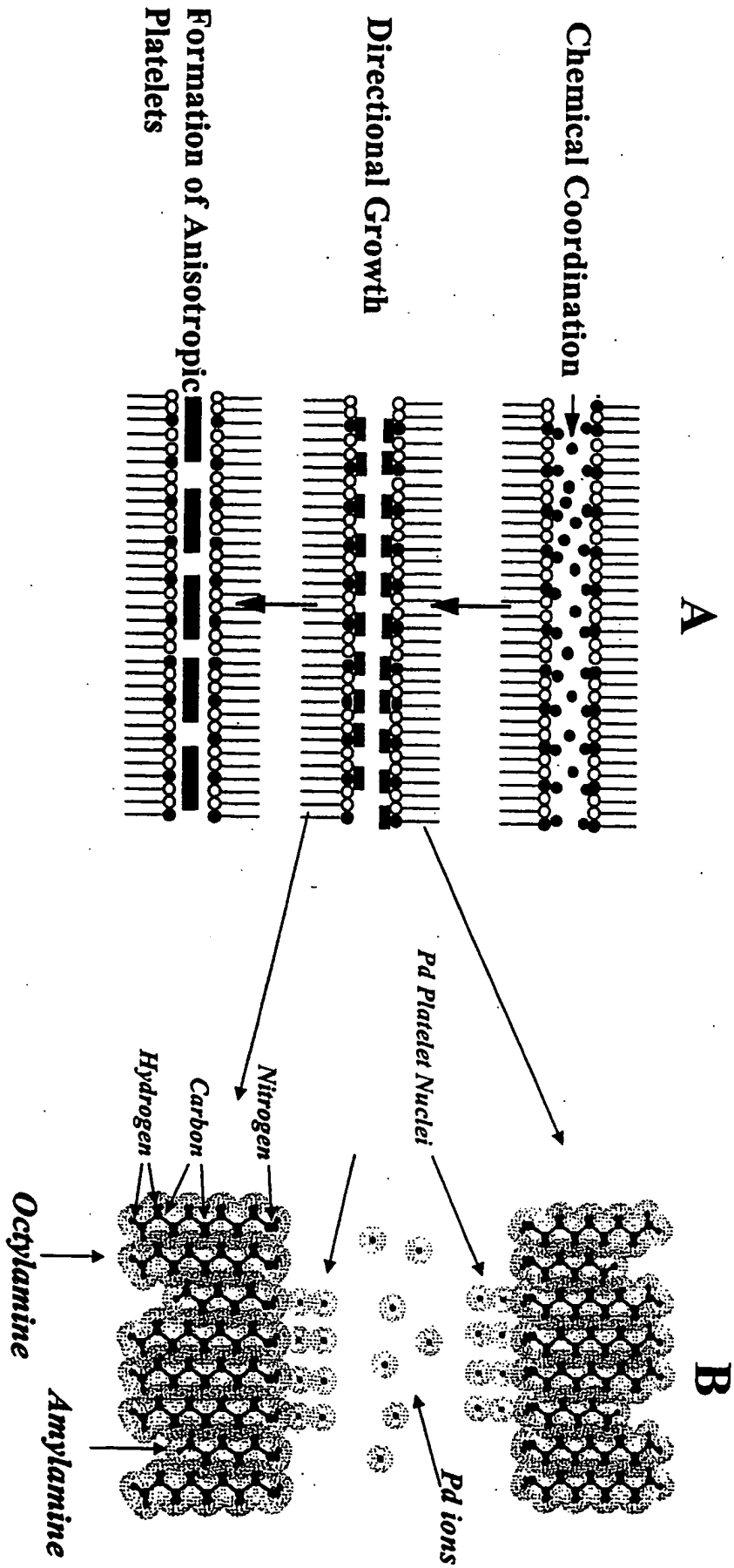


FIG. 12

FIG. 13

Synthetic Approach For Platelet Pd Particles



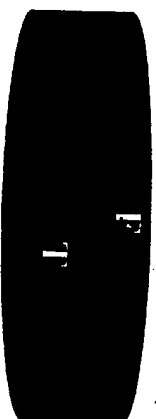
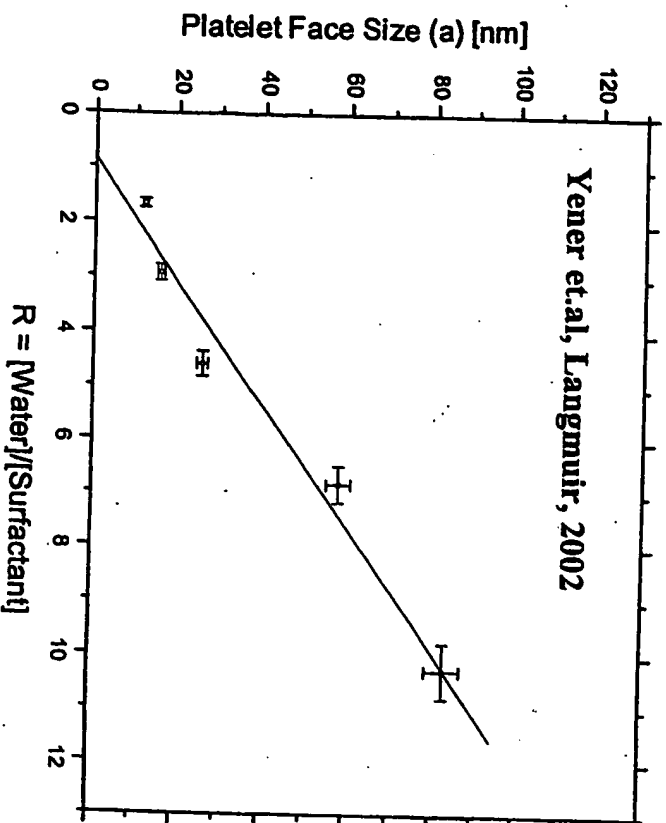
Platelet Size Control

$$R = [\text{Water}]/[\text{Surfactant}]$$

Surfactant = Octylamine + 5% Amylamine



The platelet face size (a) as a function of R.



F = Face

T = Thickness

FIG. 14

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Platelets Made

Sample	R Value	Amount Made	F*	T*
Pd 1-75	6.8	20.4mg		
Pd 1-77	20.4	228mg		
Pd 1-80-3.4	3.4	66mg		
Pd 1-80-1.7	1.7	33mg		
Pd 1-86	10.2	236mg		
Pd 1-91-8.5	8.5	212mg		
Pd 1-91-6.8	6.8	254mg		
Pd 2-5-1.7	1.7	21.2		
Pd 2-5-3.4	3.4	21.2		
Pd 2-5-6.8	6.8	21.2	33nm	2nm
Pd 2-5-8.5	8.5	21.2		
Pd 2-5-10.2	1.2	21.2	150nm	6nm
Pd 2-5-20.4	20.4	21.2		
Pd 2-8a	10.2	21.2	130nm	8nm
Pd 2-8b (AA)	10.2	21.2	80nm	1.7nm

*



FIG. 15

nMg Materials Synthesized

Sample	Amount Made
Mg 1-2	0.198g
Mg 1-10	0.4g
Mg 1-13	0.4g
Mg 1-18	0.2g
Mg 1-41	0.2g
Mg 1-51	0.4g
Mg 1-57a	0.5g
Mg 1-57b	0.5g
Mg 1-57c	0.5g

Platelet nPd

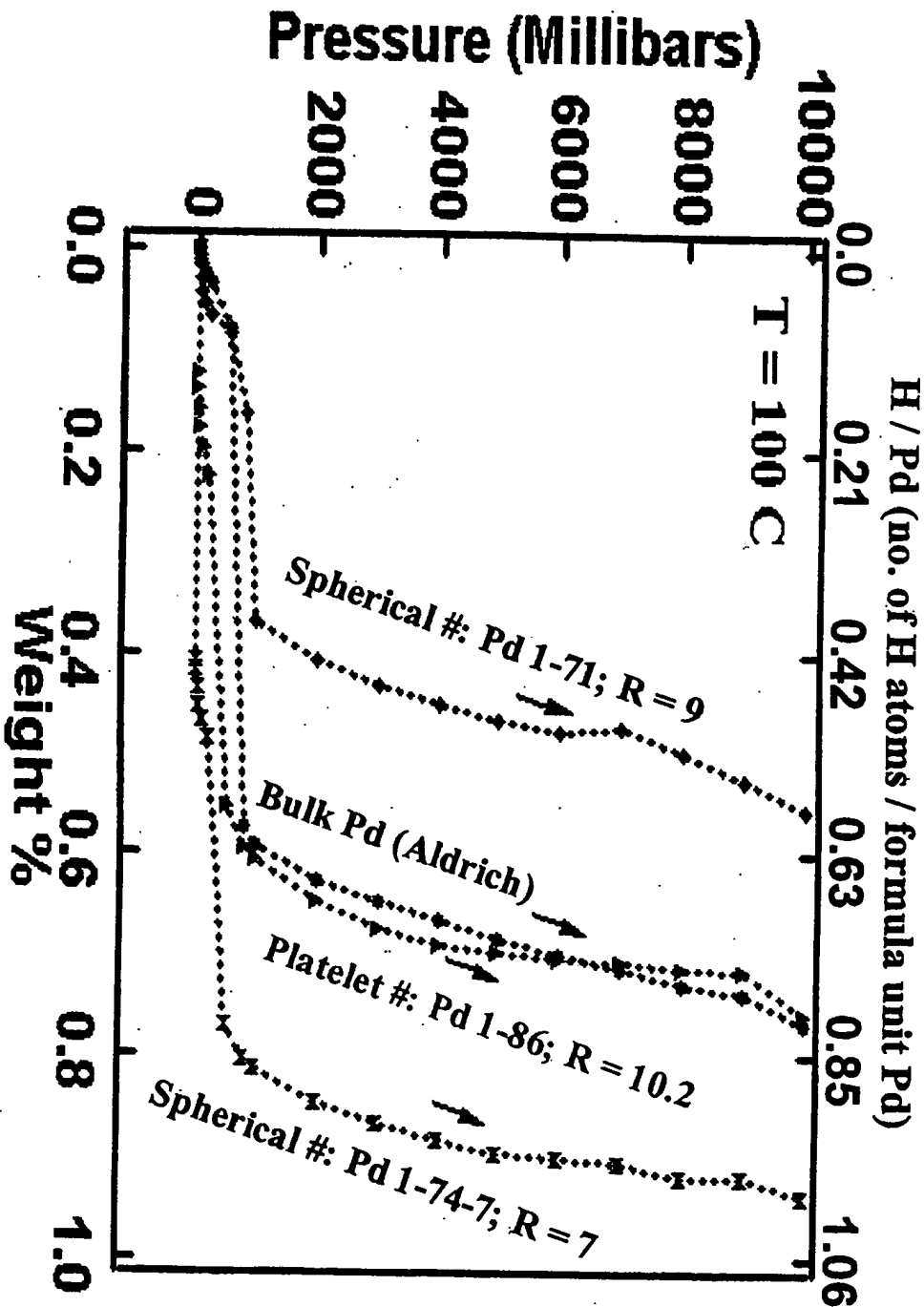
Sample	R Value	Amount Made
Pd 1-75	6.8	20.4mg
Pd 1-77	20.4	228mg
Pd 1-80-3.4	3.4	66mg
Pd 1-80-1.7	1.7	33mg
Pd 1-86	10.2	236mg
Pd 1-91-8.5	8.5	212mg
Pd 1-91-6.8	6.8	254mg
Pd 2-8-1.7	1.7	21.2
Pd 2-8-3.4	3.4	21.2
Pd 2-8-6.8	6.8	21.2
Pd 2-8-8.5	8.5	21.2
Pd 2-8-10.2	1.2	21.2
Pd 2-8-20.4	20.4	21.2
Pd 2-8a	10.2	21.2
Pd 2-8b (AA)	10.2	21.2

Spherical nPd

Sample	R Value	Amount Made
Pd 1-62	8	1.3mg
Pd 1-67	6	4mg
Pd 1-68-1	1	2mg
Pd 1-68-2	2	4mg
Pd 1-69-1	1	.67mg
Pd 1-68-2	2	1.35mg
Pd 1-69-6	6	12.3mg
Pd 1-69-8	8	16.32mg
Pd 1-70-7	7	19mg
Pd 1-70-9	9	30.5mg
Pd 1-71	9	61mg
Pd 1-73	5	63.6mg
Pd 1-74-3	3	61mg
Pd 1-74-7	7	71mg
Pd 1-91-6	6	81.4mg
Pd 1-91-8	8	108.5mg
Pd 1-96	7	399mg
Pd 1-100	25	84.8mg

FIG. 16

Adsorption Isotherms of Palladium NanoParticles and Bulk Sample at 100 °C



Bulk sample particle size: 1.0 – 1.5 Microns
 Spherical particles: R = 7 (5nm) and R = 9(8nm);
 Platelets R=10.2 (8nm thick)

FIG. 17

Comparison of TGA Evaluations at 100C for Nano and Bulk Samples

Sample	Knee for H/Pd Ratio	Plateau Onset (Wt %)	Estimated Adsorption Rate* (H/Pd/min)	Estimated Desorption Rate* (H/Pd/min)
Pd 1-74-7 R = 7 (4-5 nm Spheres)	~ 0.78	~ 0.47	~0.01	~0.005
Pd 1-86 R = 10.2 (8 nm thick Platelets)	~ 0.65	~ 0.2	0.007	0.003
Bulk (1.0 – 1.5µm)	~ 0.6	~ 0.08	0.005	0.002

* Possibly affected by sintering

FIG. 18

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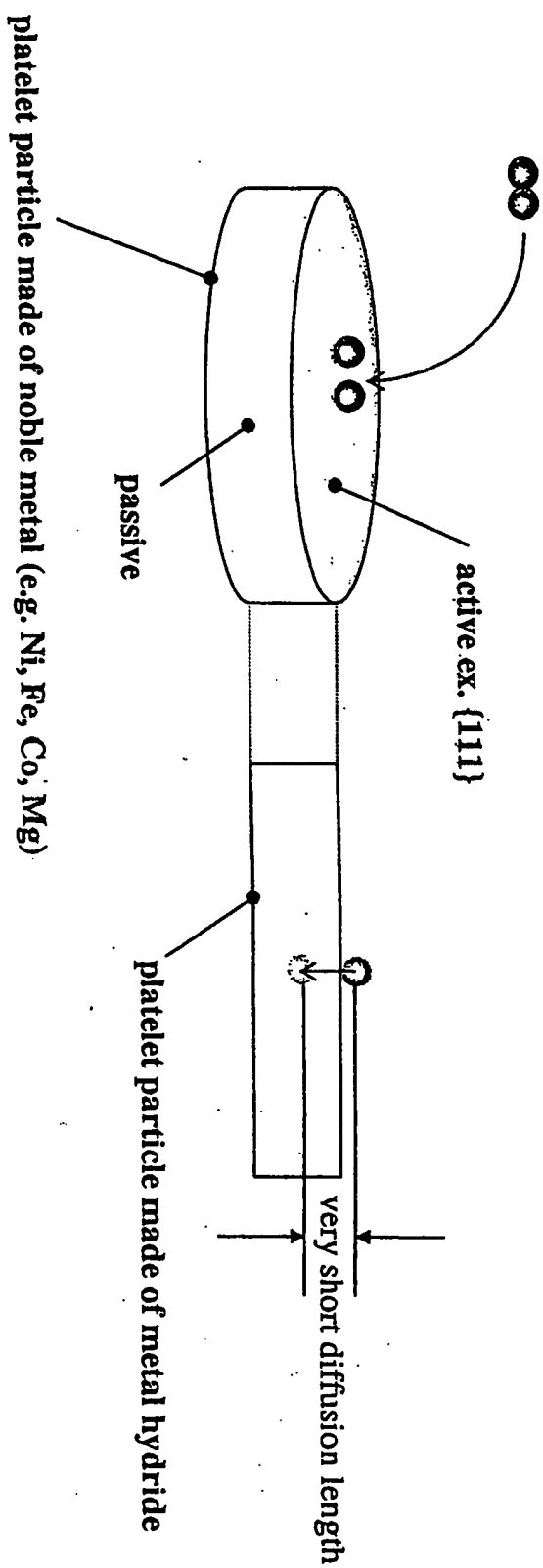


FIG. 19

FIG. 20

FIG. 21 TGA Compositional Analysis

Sample	Pd in sample	Wt.%Pd as PdO in sample	Wt.% Carbon	η corr. factor
Spherical				
Pd 2-35 R=2 (4nm)	95.34%	0%	2.37%	1.05
Pd 2-71 R=8	96.16%	0%	0.05%	1.04
Platelet				
Pd 2-65-10 R=10 (327nm X 2.8nm to 109nm X 1.5nm)	87.26%	0%	0.08%	1.15
Pd 2-65-7 R=7 (82nm X 1.3nm)	94.69%	0%	0.17%	1.06

FIG. 22 NanoCrystalline vs NanoParticle

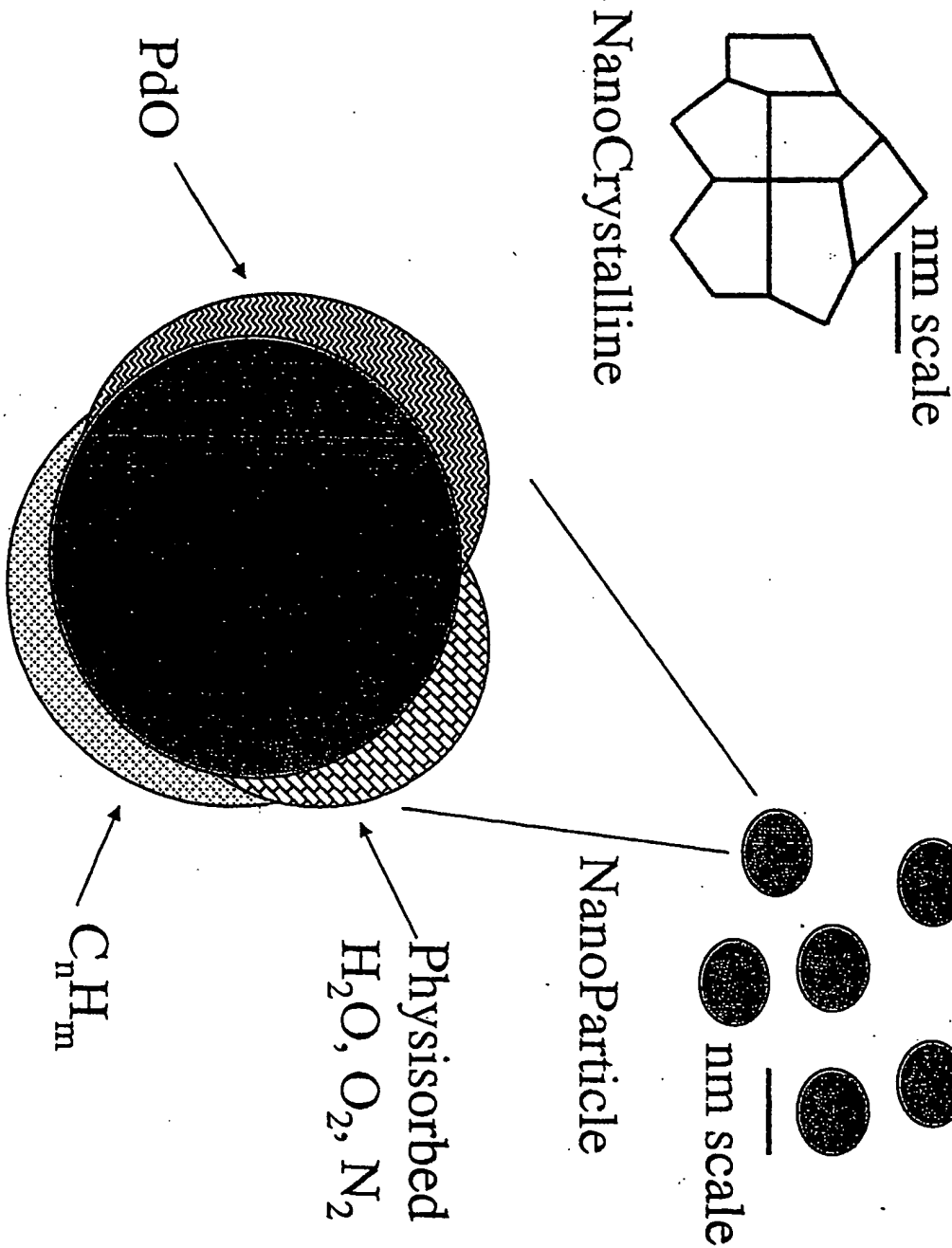
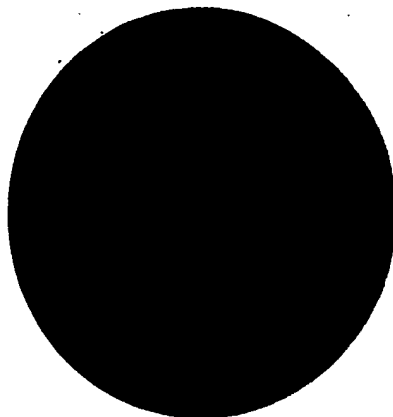


FIG. 23 . Effect of morphology on
 particle packing

Spherical



$r \sim 3-50 \text{ nm}$

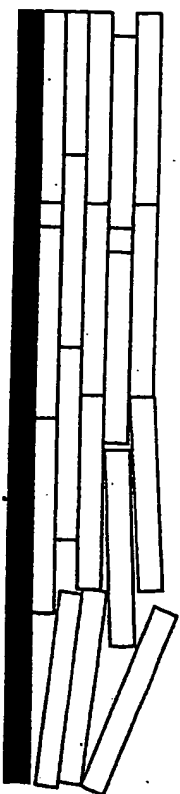
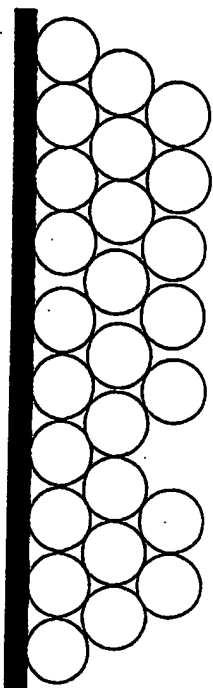
Platelet

VS



$t \sim 1-200 \text{ nm}$
 $a/t \sim 10-100$

Particle Packing Characteristics



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FIG. 24 HRTEM of Platelet nanopd



Moire fringes

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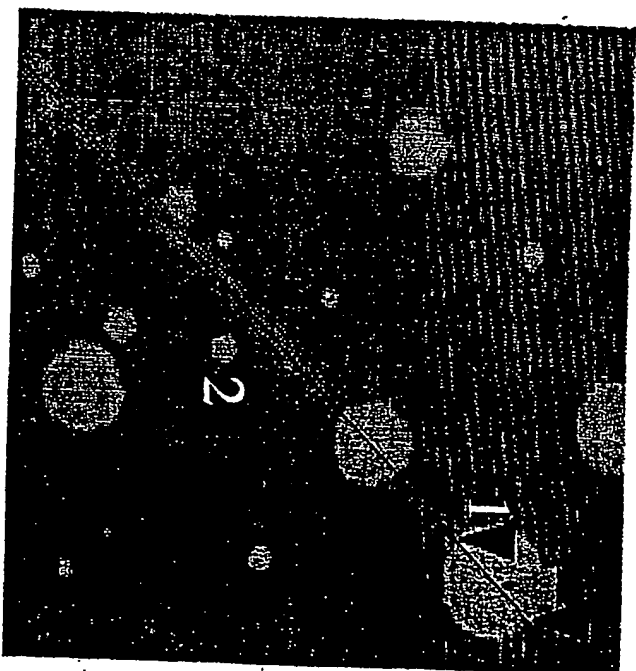
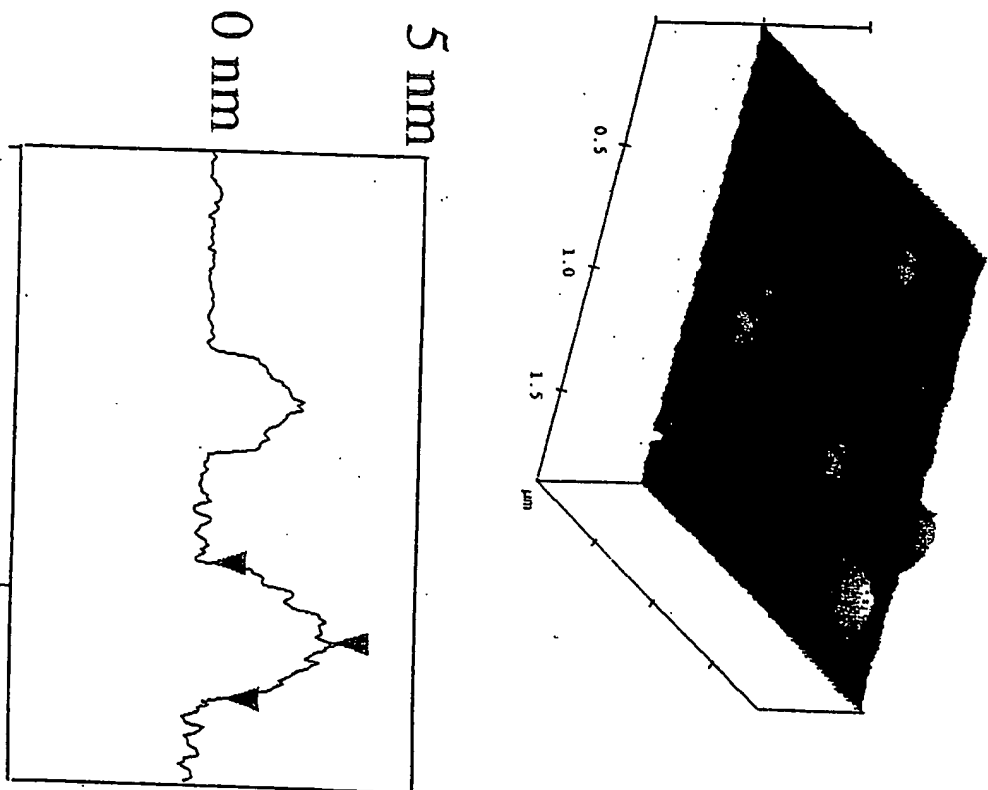
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FIG. 25
AFM of Platelet nanoPd



- (1) 327nm X 2.8nm
- (2) 109nm X 1.5nm

FIG. 26 H Adsorption of Spherical Pd

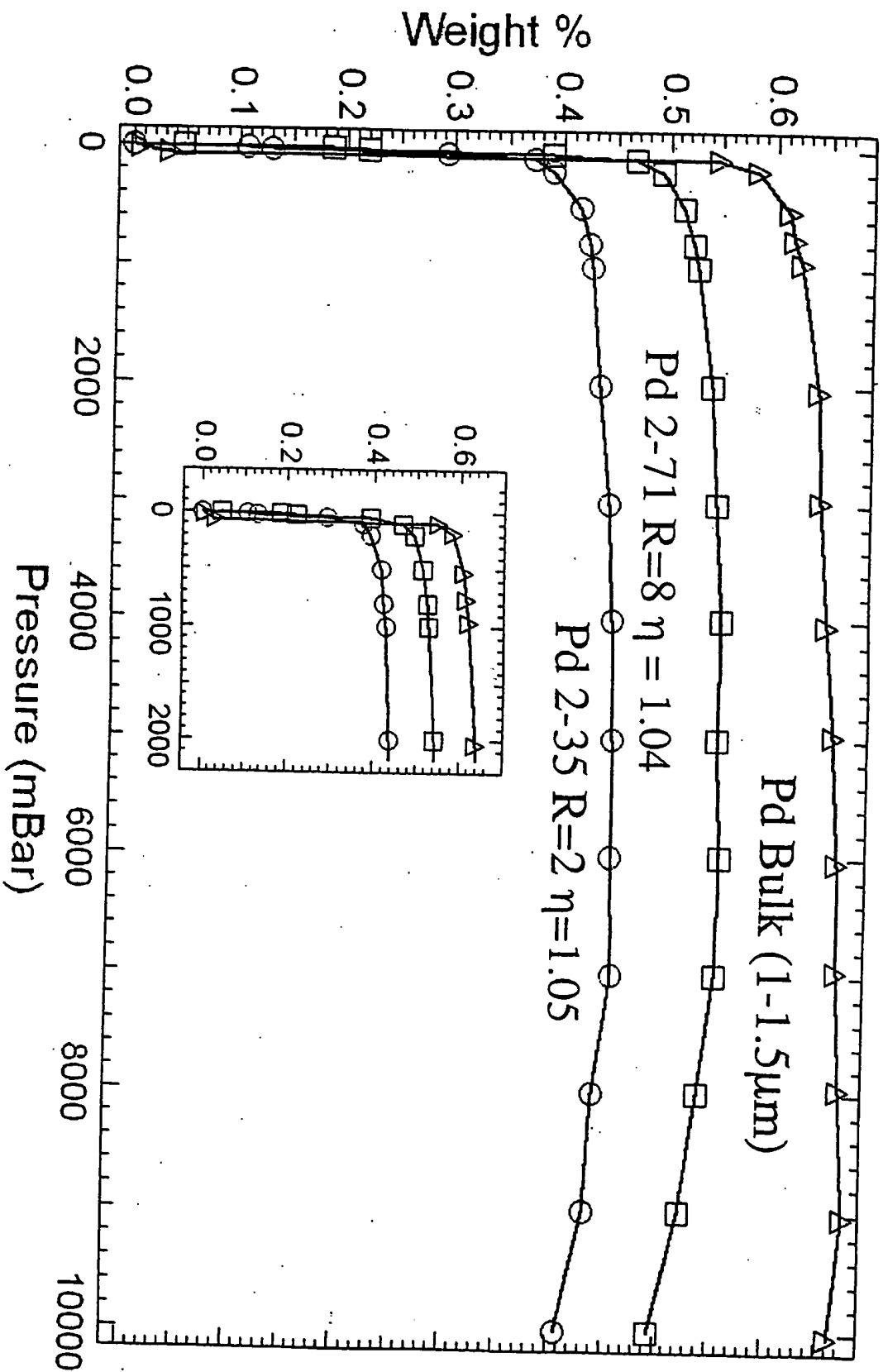


FIG. 27 H₂ Adsorption of Pd Platelets

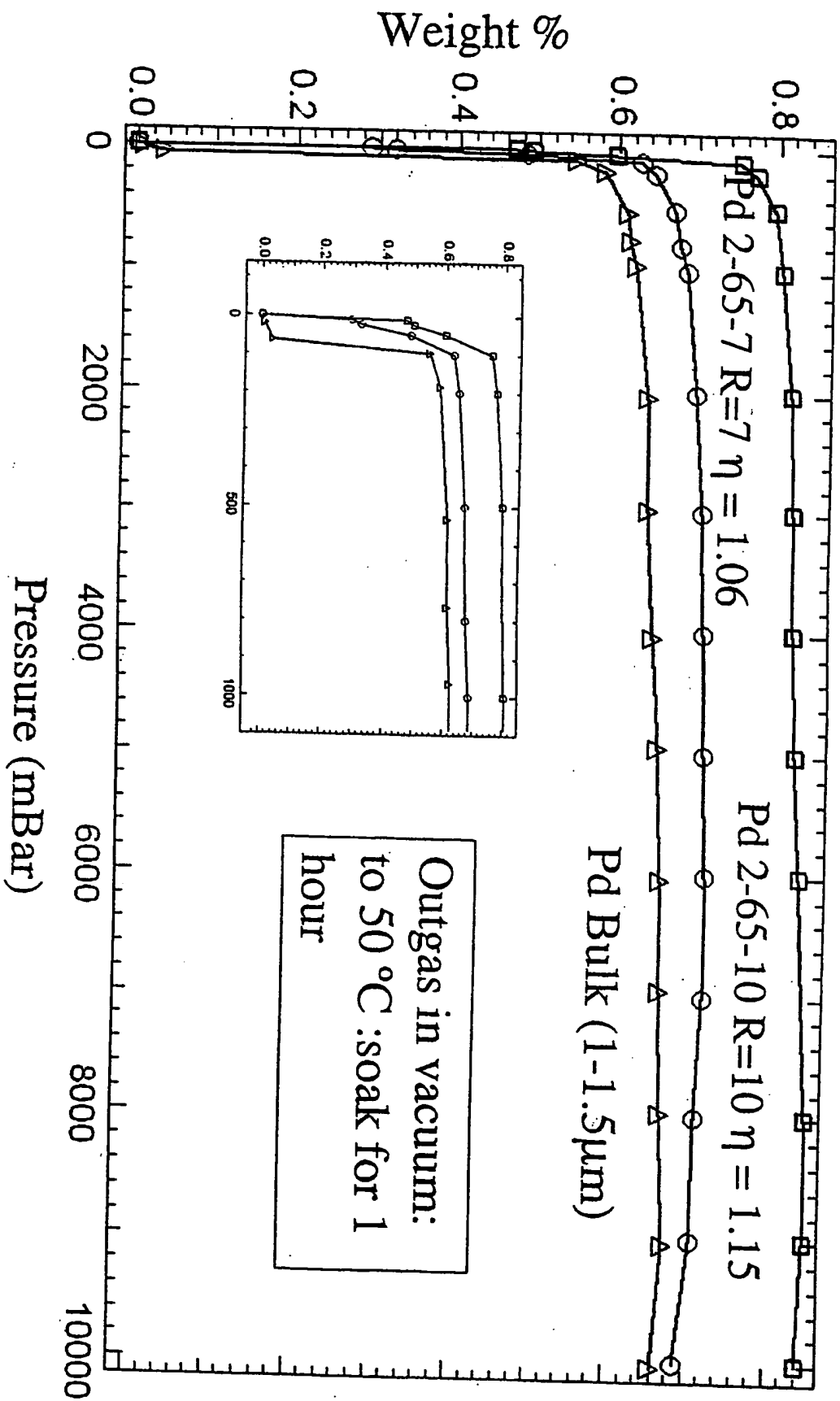
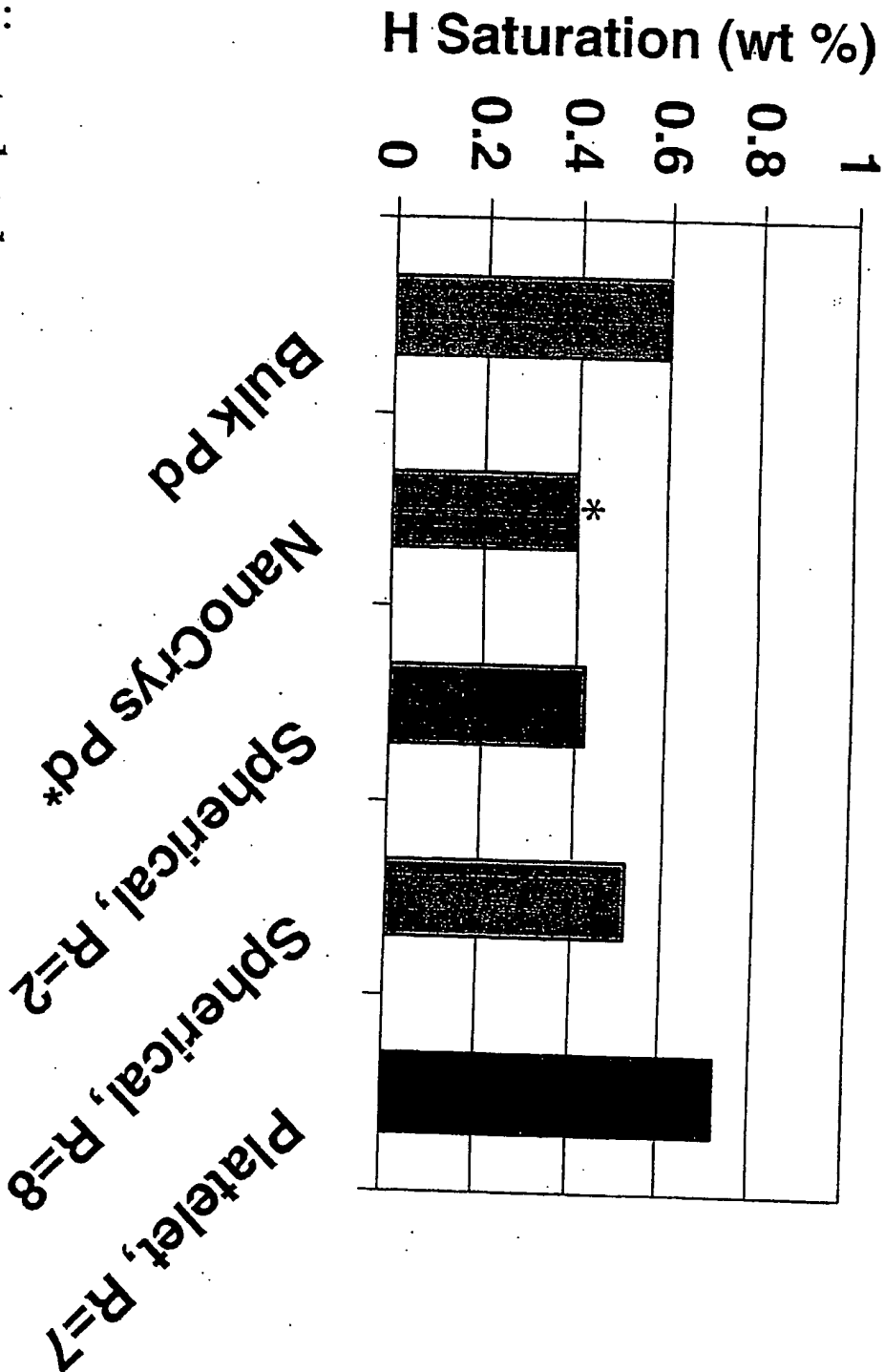


FIG. 28 Hydrogen Saturation vs Pd Material



* Kuji, et.al, taken as maximum H/Pd at 0.1 MPa at 353K

Hydrogen Absorption Isotherms of Pd Nanoparticles (through Thermogravimetric Analysis)

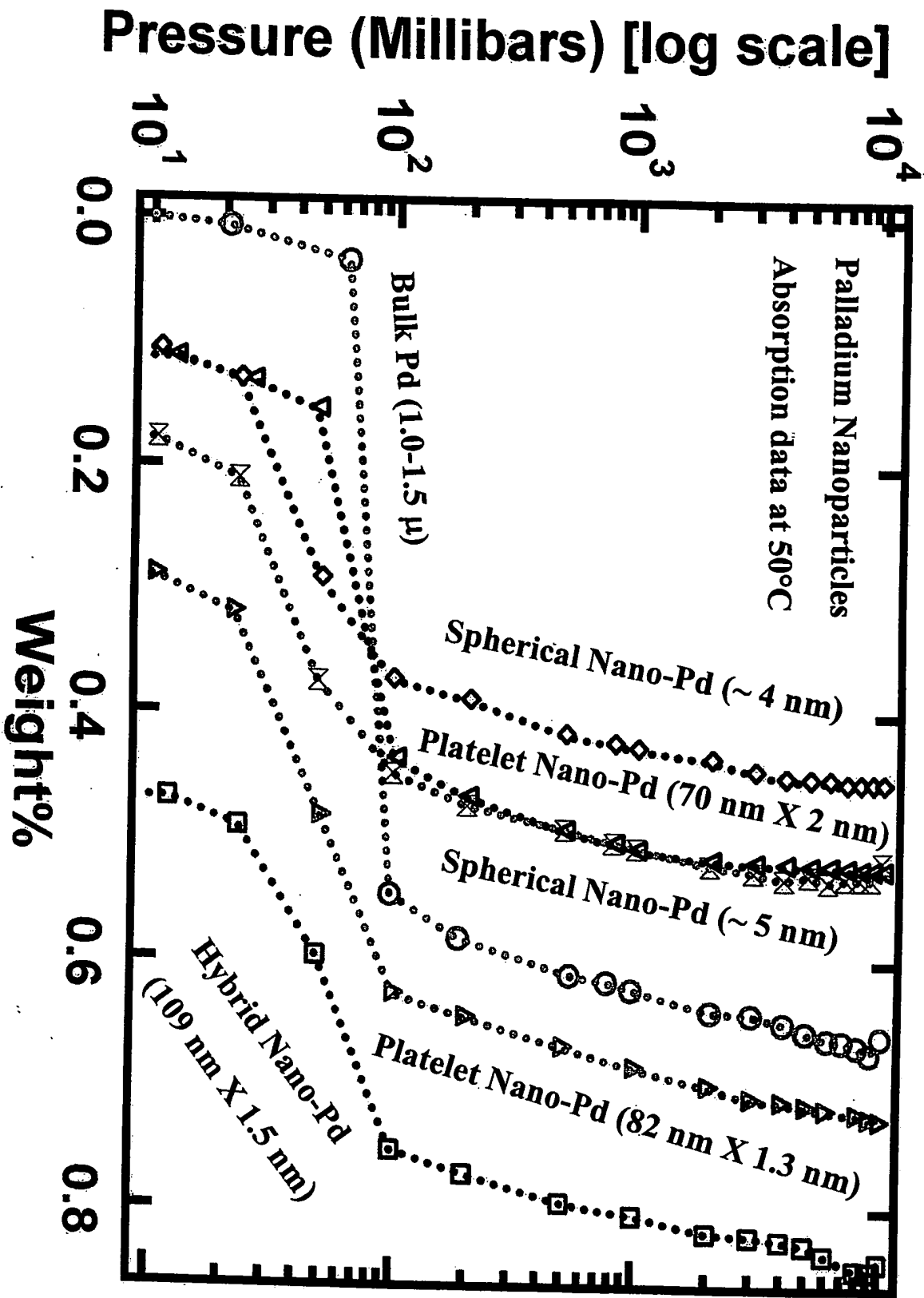


FIG. 29

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FIG. 30
HRTEM of Platelet Particles

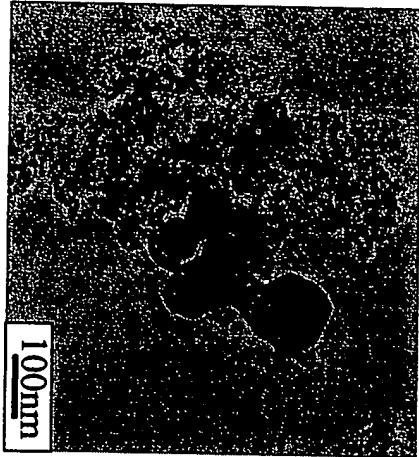
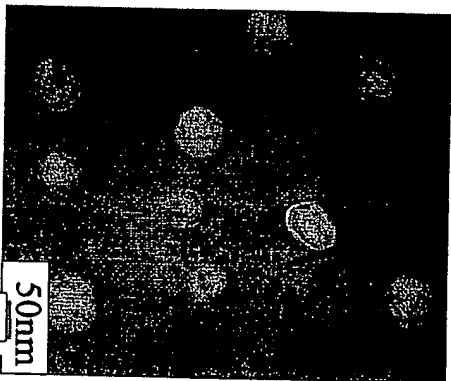
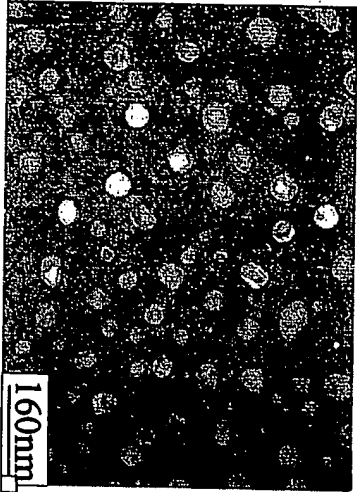


Fig 20g

Mixed (hybride) Platelet Sample

Fig 30b



Platelet Sample average size 70nm X 1.7nm

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Review of nanoPd Sample tested for Hydrogen Storage

<u>Sample</u>	<u>R</u>	<u>%Pd</u>	<u>d¹</u>	<u>a²</u>	<u>t³</u>
<u>Reverse Micelle Synthesis</u>					
2-35*	spherical	2	95	4	
2-71*	spherical	8	96	5	
2-44	spherical	5	82		
<u>Bilayer Synthesis</u>					
2-65-10	mixed	10	87	5	327 2.8
3-18	platelet	8	96		109 1.5
3-33	platelet	6	94		70 2
				130	2.6

R = [water]/[Surfactant], 1 = diameter of spherical particle, 2 = face size of platelet particle, and 3 = thickness of platelet particle in nm. * Samples washed with hydrazine

FIG. 31

Sintering of Platelet Particles

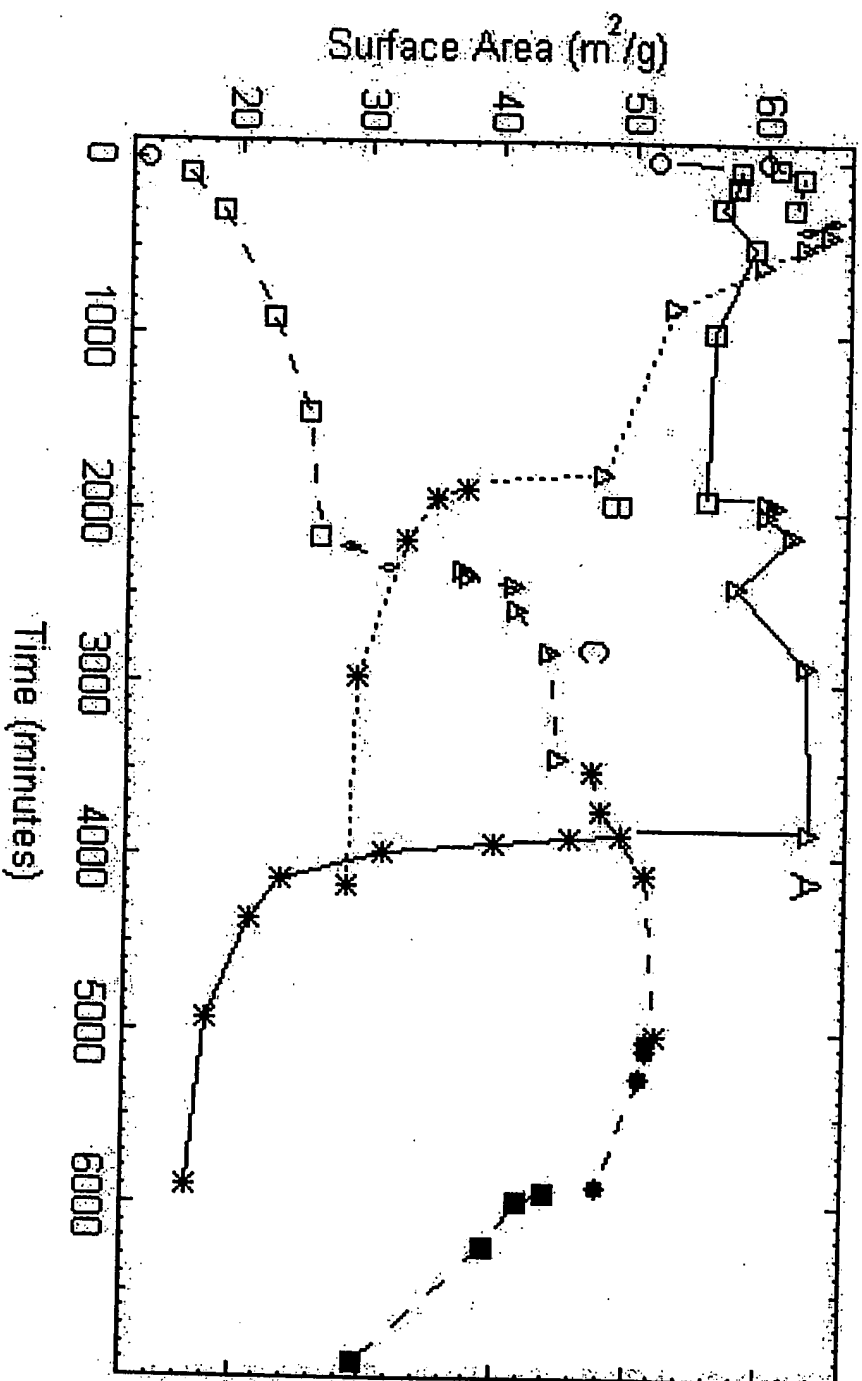
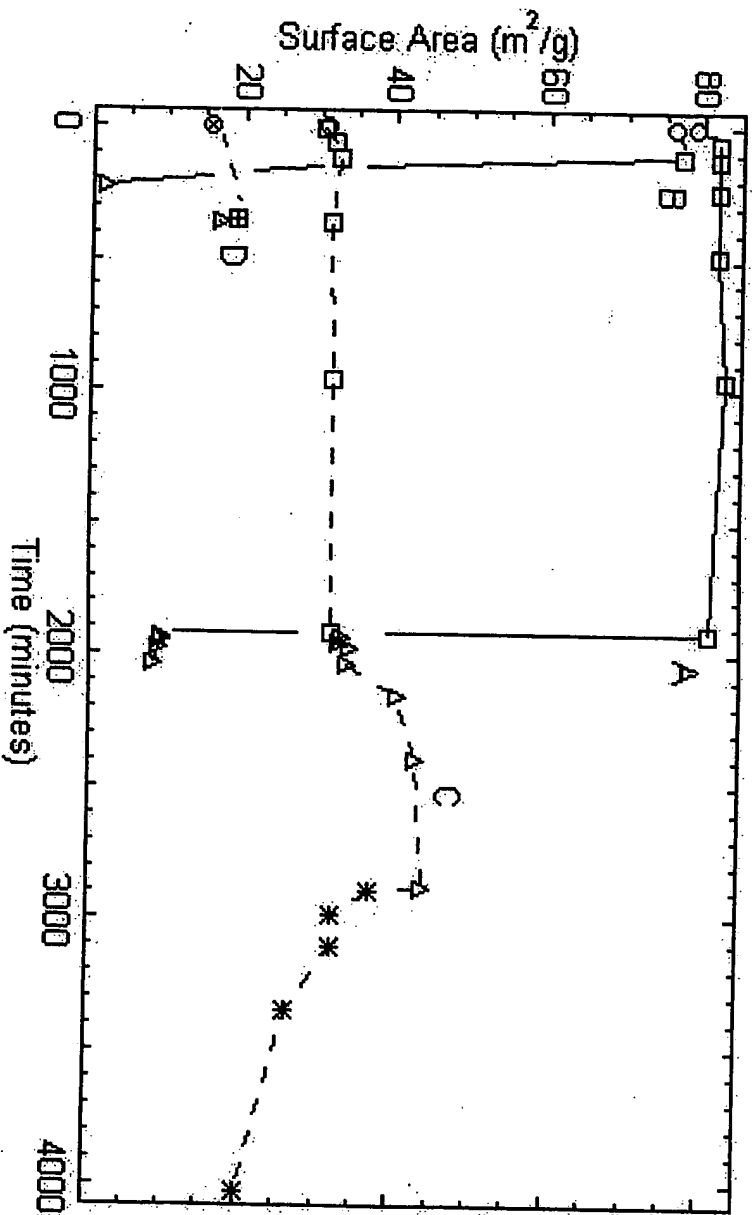


FIG. 32

Surface area vs. heating time for sample synthesized by bilayers. Samples A(sample 3-18) and B(sample 3-33) consist of only platelets. Sample C(sample 2-65-10) is a mix of platelets and spherical particles. Heating temperatures are O = room temperature, □ = 50 °C, ◇ = 75 °C, △ = 100 °C, * = 150 °C, ● = 200 °C, ■ = 250 °C.

Sintering of Spherical Particles



Surface area vs. heating time for spherical particles synthesized via reverse micelles. A(sample 2-44) and B(sample 2-35) = not washed with hydrazine hydrate. C(sample 2-71) and D(sample 2-35) = washed with hydrazine hydrate. Lines B and D are both of sample 2-35. Heating temperatures are O = room temperature, □ = 50 °C, △ = 100 °C, and * = 150 °C.

FIG. 33

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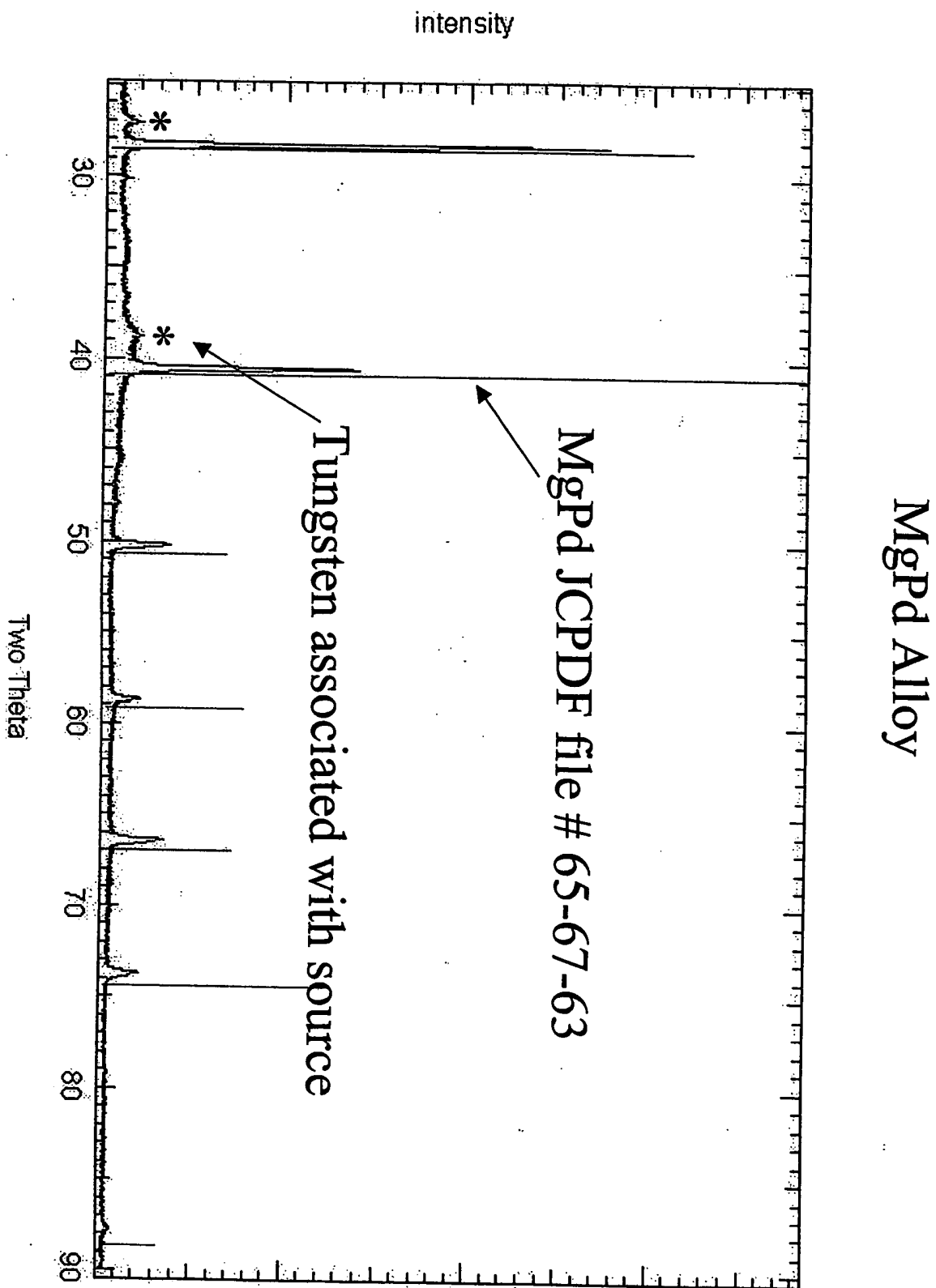
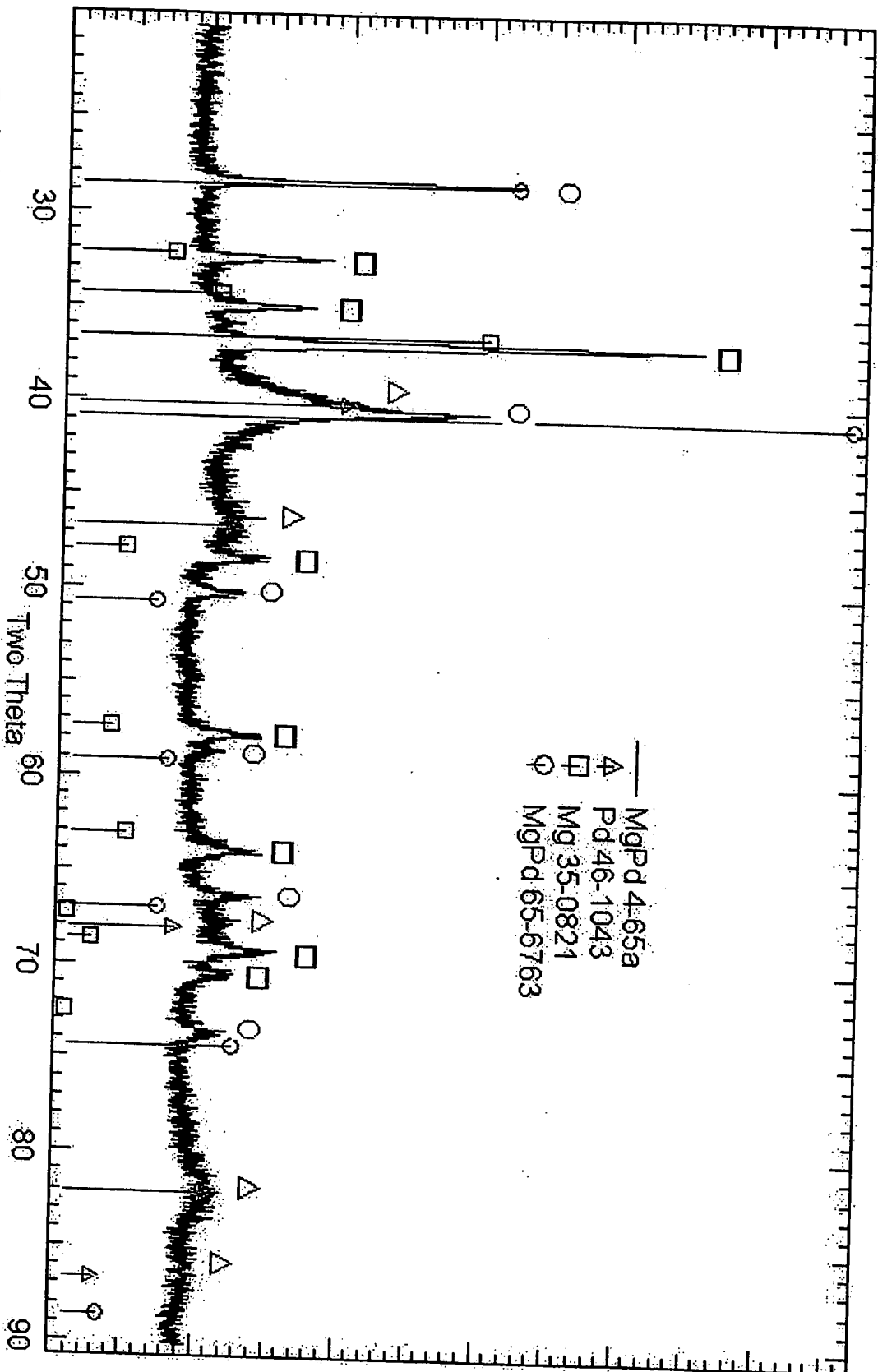


FIG. 34

MgPd Alloy with Mg and Pd Metal



It is believed that excess Mg in the reaction lead to the formation of MgPd alloy along with metallic Pd and Mg.

FIG. 35